

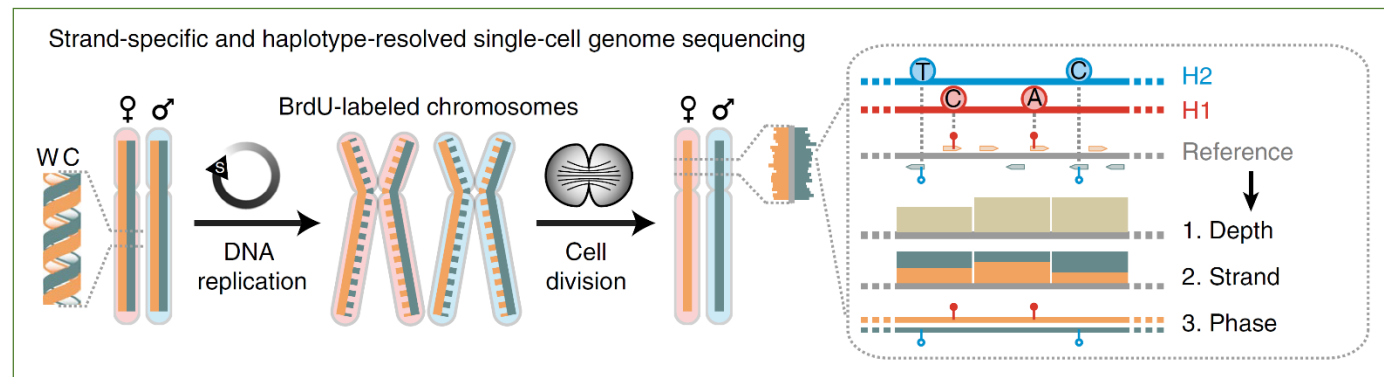
# Single Cell Tri-Channel Processing (scTRIP)

## Challenge

- Structural variation (SV), such as deletions, duplications, inversions or translocations in the DNA, is a major source of genetic variation and plays a role in numerous diseases.
- The discovery of SVs can help to better understand disease, enable precision medicine approaches, and allow for quality control in gene and cell therapy approaches.
- However, studying SVs is limited by high costs for sequencing coverage, challenges to discover SVs in repetitive regions and lacking approaches for single cell SV tracking.

## Technology

Strand-specific single cell sequencing data are analyzed by a computational, bayesian framework to integrate three layers of information (depth of coverage, read orientation and haplotype phase) for calling SVs and to derive a specific 'diagnostic footprint'.



Haplotype-aware discovery of SVs in single cells by scTRIP: Overview of the Strand-seq protocol used to preserve strand orientation and homolog (haplotype) identity (for details see [Sanders et al., 2019](#)). Right: Haplotagging approach assigning individual Strand-seq reads to either haplotype (H)1 or H2. Red lollipops mark reads assigned to H1 based on overlapping SNPs; blue lollipops mark H2 reads. From this, three data layers are considered: (1) the total number of reads in a binned region are measured to calculate the 'depth' layer; (2) the relative proportion of W and C reads are measured to calculate the 'strand' layer; and (3) the number of W and C reads assigned to H1 or H2 are used to calculate the 'phase' layer.

### Internal EMBLEM Reference

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# Single Cell Tri-Channel Processing (scTRIP)

## Advantages

- Systematic, detailed and accurate
- Includes all known forms of karyotypic abnormalities
- Strand-specific single cell sequencing
- Integration of multiple information layers
- Less required sequencing depth than other methods
- More reliable than current standard
- cost-effective

## Applications

- Disease classification for precision medicine
- Patient stratification for clinical studies
- Assessment of genome integrity in cell and gene therapy applications
- Therapeutic CRISPR-Cas9 editing
- Rare disease genetics.

## Intellectual Property

A patent application has been filed.

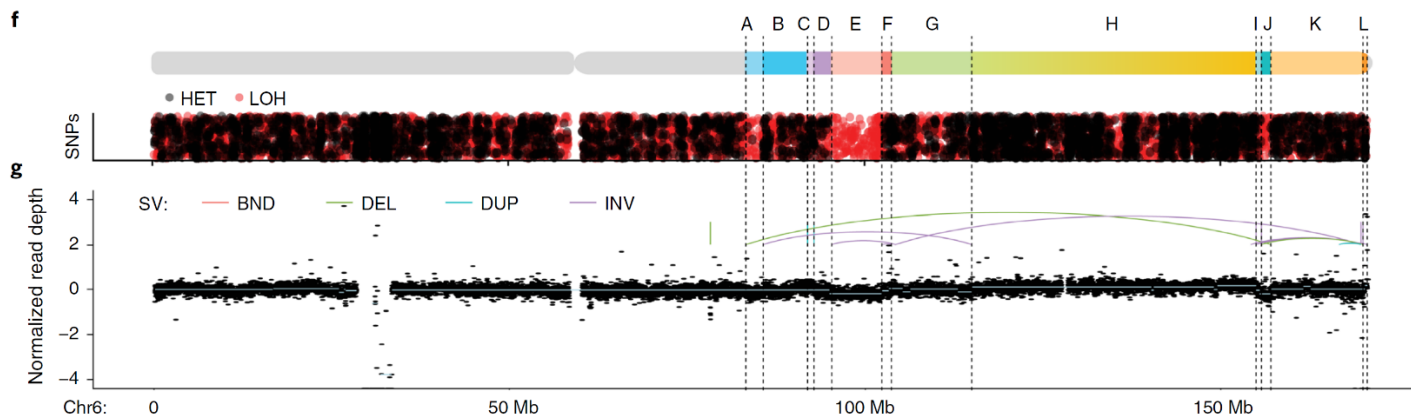
## Commercial Opportunity

The technology is ready to use. We offer a technology evaluation program as well as a licensing or collaboration/co-development opportunity.

## Further Reading

[EMBL News](#)

[Sanders et al., 2019](#)



**Application example:** Locating previously unrecognized SVs in a T-cell acute lymphoblastic leukemia (T-ALL) relapse sample (for details see [Sanders et al., 2019](#)).

**f**, Detection of interspersed losses and retention of LOH in conjunction with the clustered SVs, indicative for a DNA rearrangement burst<sup>43</sup>. Regions with normal density of reference heterozygous SNPs (red), but with decreased density of heterozygous SNPs detected in P33 (black), are indicative for LOH (see Methods).

**g**, Verification of subclonal clustered rearrangement burst at 6q. Breakpoints inferred by scTRIP are shown as dotted lines and scTRIP-inferred segments are denoted using the letters A to L. Colored breakpoint-connecting lines depict the paired-end-mapping-based rearrangement graph (BND: translocation-type, DEL: deletion-type, DUP: tandem-duplication-type and INV: inversion-type paired-end SV predictions). Using bulk whole-exome and mate-pair sequencing, read-depth shifts at these breakpoints were subtle and thus, this subclonal complex rearrangement escaped previous de novo SV detection efforts in bulk.